

# The association of lncRNA *H19* rs217727 polymorphism with oral lichen planus in Shiraz, Iran

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## INTRODUCTION

Lichen planus (LP) is a mucocutaneous pathology that predominantly affects females in their mid-life.

**Background:** This study aimed to assess the single nucleotide polymorphisms (SNPs) of lncRNA (long non-coding RNA) *H19* rs217727 in patients with oral lichen planus (OLP) compared to controls.

**Methods:** We collected 270 DNA samples of OLP cases and healthy individuals. We used the ARMS-PCR tetra primer for DNA genotyping and applied specific primer pairs.

**Results:** The prevalence of the rs217727 C allele was lower in OLP cases than in healthy subjects ( $P = 0.005$ ). The prevalence of TT genotypes of *H19* rs217727 was greater in OLP patients compared with healthy subjects (5.71% vs 1.5%). Also, the TT genotype in the codominant model was associated with a 5.15-fold higher risk of OLP ( $P = 0.02$ ). In the dominant model, the CT+TT genotypes were associated with a 2.09-fold greater risk of OLP ( $P = 0.007$ ). The *H19* rs217727 polymorphism was linked to a 3.95-fold greater risk of OLP in the recessive model ( $P = 0.05$ ) (TT vs. CC+CT). Also, in the over-dominant model, the CT genotypes were related to a 1.78-fold greater risk of OLP ( $P = 0.03$ ).

**Conclusion:** This study demonstrated a significant link between lncRNA *H19* polymorphism and OLP lesions. Further studies on larger populations are necessary to confirm this relationship.

**Keywords:** *H19* long non-coding RNA, oral lichen planus, genetics

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The disease involves all populations; no racial groups have been recognized as more susceptible <sup>1,2</sup>. Oral lichen planus (OLP) is an inflammatory disease with

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a chronic nature and undetermined etiology affecting the oral mucosa. This non-communicable oral lesion affects more than 1-2% of the population in the 4<sup>th</sup> to 5<sup>th</sup> decade of life. OLP has various forms including white striae (reticular), white papules (papular), plaque-like, erythematous/atrophic, erosive, or bullous. The buccal mucosa is the most affected site in the oral cavity, but it may involve different parts of the mucosa and can be seen as severe erythema throughout the gingiva with or without genital or skin lesions<sup>1,3-6</sup>.

As a possibly cancerous lesion, OLP is associated with a higher risk of oral squamous cell carcinoma (OSCC). Malignant change is expected to arise in 0.5-3% of cases, though no predictive indicators distinguish the forms with an elevated risk. Hence, every patient should be checked thoroughly to identify initial cancerous potentials<sup>7-10</sup>.

Long non-coding RNAs (lncRNAs) are RNA particles extending from 200 nt to > 100 kb in size. Considering transcriptional profiling, lncRNA expression varies across human cancers<sup>11</sup>. Cell variation, gene control, chromatin renovation, malignant cell assault, and metastatic tumor development are the main actions performed by lncRNAs. Various surveys have shown the significance of lncRNAs in identifying and predicting many diseases and malignancies<sup>12-15</sup>.

As the earliest detected lncRNA, the lncRNA *H19* gene is found on chromosome 11p15.5, coding a polyadenylated, intertwined, and excelled non-coding RNA<sup>16</sup>. lncRNA *H19* is copied from the maternal hereditary alleles and is accountable for managing genome representation at distinct degrees<sup>17</sup>. *H19* can deactivate tumor suppressor proteins by interacting with p53, equally controlling the roles of oncogenes and tumor suppressor genes<sup>18,19</sup>.

*H19* polymorphism is, according to several distinct surveys, correlated with the presence and growth of tumors<sup>20,21,22</sup>. According to many types of research, single-nucleotide polymorphisms (SNPs) are critical indicators relating phenotypic variations to DNA-sequence modifications<sup>19</sup>. Investigations in this area are anticipated to explain the structural biology and clarify the molecular origin of disorders. Despite the significance of lncRNA *H19* in numerous diseases, its correlation with OLP is yet to be assessed. Hence, we examined the link between susceptibility to OLP

and SNPs in the lncRNA *H19* rs217727 gene in an Iranian population.

## METHODS

### Patients

This cross-sectional study involved 270 subjects recruited between 2018–2019. We had two groups: OLP patients as the case group and healthy subjects as the control group. The patients were examined intraorally by an oral medicine specialist. Cases with clinical signs and histopathological confirmation of OLP were evaluated. Subjects with a history of autoimmune pathology, malignancy, chemotherapy, radiotherapy, or patients using medications related to lichenoid reactions in the past six months and lichenoid contact reactions were excluded from this study. Subjects referred to Shiraz Dental School for routine dental examination were chosen as the control group; they were non-smokers, and all exclusion criteria mentioned for the cases were also considered for controls.

All research participants were Iranian subjects who had no kinship with one another. Patient information was collected in person. All patients participated voluntarily and signed an informed consent. A trained nurse obtained 5 mL of peripheral blood from each participant. The Ethics Committee of Shiraz University of Medical Sciences approved the research protocol under the ethics code IR.SUMS.DENTAL.REC.1399.199.

### Single nucleotide polymorphism (SNP) selection

Using the dbSNP database, the region of *H19* and its promoter were selected. A Hardy-Weinberg equilibrium (HWE) value  $\geq 0.05$  for the Iranian population was selected using Haploview software; finally, the polymorphic site (rs217727) was selected.

### Genotyping

DNA was extracted using the salting-out technique. SNP genotyping was carried out using specific primers via the tetra-primer (ARMS-PCR) method (Table 1). The 25  $\mu$ L mixture for the PCR reaction included 50–100ng of DNA, 12.5  $\mu$ L of Taq DNA polymerase (Taq Pol) II Master mix Red (Amplicon, Denmark), 0.5  $\mu$ L of inner and 0.25 $\mu$ L of outer primer pairs (10 pm/ $\mu$ L), and distilled deionized water to reach 25  $\mu$ L.

**Table 1.** Primers and rs217727 tetra-primer ARMS-PCR product sizes.

Primer name	Oligo sequence 5'--> 3'	Product size
OutF1	ATGACTCAGGAATCGGCTCTGGAAGGTG	Product size of two outer primers: 397 bp
OutR1	GGGAAACAGAGTCGTGGAGGCTTTGA	
inF1	TCATCTTCATGGCCACCCCTGCTGT	Product size for C allele: 248 bp
inR1	ATATGGTGGCTGGTGGTCAACCGTACG	Product size for T allele: 200 bp

The PCR conditions included an initial denaturing cycle at 95 °C for 5 minutes, after that 30-second cycles of denaturing at 95 °C for 30 seconds, annealing at 65 °C for 30 seconds, and extension at 72 °C for 30 seconds, and a final extension at 72 °C for 5 minutes. Negative controls—tubes containing the PCR mixture without the DNA template—were included in every run to examine for contamination. After that, PCR products were electrophoresed on 3% agarose gel. The genotyping data was checked by conducting conventional PCR in a reaction mixture of 50 µL using primer pairs OutF1 and OutR1 with the abovementioned PCR conditions<sup>20</sup>. After that, Sanger sequencing was performed. The schematic presentation of the tetra-primer ARMS-PCR technique is shown in Figure 1. This methodology was similar

to our previous studies on patients with squamous cell carcinoma (SCC) or salivary gland tumors<sup>20,21</sup>.

### Statistical analysis

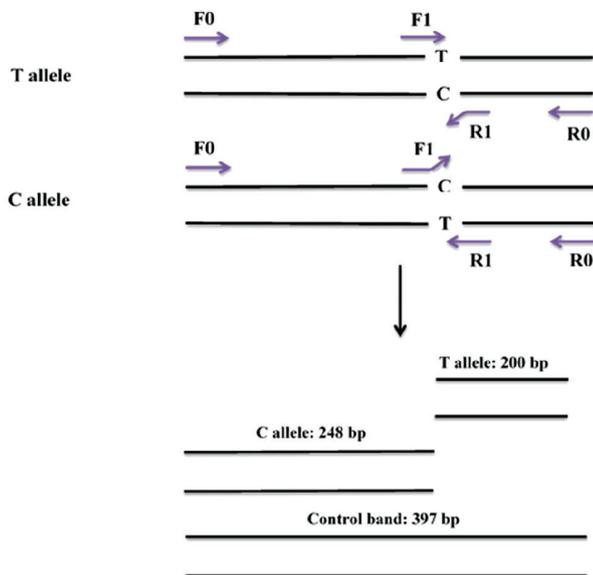
To evaluate the genotype frequencies among the controls, the HWE was checked by the chi-squared ( $\chi^2$ ) test. The association of *H19* rs217727 genotypes with risk of OLP was calculated by odds ratio (OR) and 95 percent confidence intervals (CI) by logistic regression analysis. In all tests,  $P < 0.05$  was regarded as significant.

### RESULTS

The OLP patients included 46 females (66%) and 24 males (34%), with an age range of 18–70 and a mean age of  $46.4 \pm 12.2$  years. The control group included 45 females (22.5%) and 155 males (77.5%), with a mean age of  $58.79 \pm 17.14$  (18–85) years.

The product sizes in the PCR test of *H19* rs217727 polymorphism were 248 bp for the C allele, 200 bp for the T allele, and 397 bp for the internal control with 3% agarose gel (Table 1).

The prevalence of the rs217727 SNP genotypes and their correlations with OLP risk are demonstrated in Table 2. The findings illustrate an association between OLP and rs217727 polymorphism in the Iranian population. The prevalence of the rs217727 C allele was lower in OLP patients compared to healthy subjects (C vs. T: OR = 0.53, 95% CI: 0.33–0.83,  $P = 0.005$ ). There was a relationship between rs217727 C allele presentation and a reduced risk of OLP ( $P = 0.005$ ). The prevalence of the TT genotype of *H19* rs217727 was significantly greater in OLP patients compared to healthy subjects (5.71% vs. 1.5%); the TT genotype was associated with a 5.15-fold greater risk of OLP in the codominant model (OR = 5.15, 95% CI = 1.01–28.76,  $P = 0.02$ ), while the CT+TT genotypes were related to a 2.09-fold higher risk of OLP in the dominant model (OR = 2.09, 95% CI = 1.20–3.66,  $P = 0.007$ ). Moreover, the *H19* rs217727 polymorphism was associated with a 3.95-



**Figure 1.** A schematic presentation of the tetra-primer ARMS-PCR method for detecting C/T single-nucleotide polymorphisms (SNPs). Two pairs of primers are used to generate two allele-specific products: one pair (F1 and R0) produced an amplicon representing the C allele, and the other pair (F0 and R1) produced an amplicon representing the T allele. The two outer primers (F0 and R0) were positioned at different distances from the polymorphic nucleotide, so the two allele-specific amplicons differed in length. This allowed them to be discriminated by gel electrophoresis.

**Table 2.** Allele and genotype frequencies of *H19* rs217727 in patients with oral lichen planus and controls.

Inheritance model	rs217727 polymorphism	Patients (&)	Healthy subjects (&)	Odd ratio (95% Confidence Interval)	P-value
Codominant <sup>a</sup>	Genotype				
	CC	34 (48.57)	133 (66.5)	1	
	CT	32 (45.71)	64 (32)	1.95 (1.10-3.45)	0.01
Dominant <sup>b</sup>	TT	4 (5.71)	3 (1.5)	5.15 (1.01-28.76)	0.02
	CC	34 (48.57)	133 (66.5)	1	
Recessive <sup>c</sup>	CT+TT	36 (51.42)	67 (33.5)	2.09 (1.20-3.66)	0.007
	CC+CT	66 (94.28)	197 (98.5)	1	
Over-dominant <sup>d</sup>	TT	4 (5.71)	3 (1.5)	3.95 (0.79-21.69)	0.05
	CC+TT	38 (54.28)	136 (68)	1	
	CT	32 (45.71)	64 (32)	1.78 (1.01-3.12)	0.03
	Allele				
	T	40 (28.57)	70 (17.5)	1	
	C	100 (71.42)	330 (82.5)	0.53 (0.33-0.83)	0.005

<sup>a</sup> The co-dominant model is defined as the major allele homozygotes vs. heterozygotes.

<sup>b</sup> The dominant model is defined as the major allele homozygotes vs. heterozygotes + minor allele homozygotes.

<sup>c</sup> The recessive model is defined as the major allele homozygotes + heterozygotes vs. minor allele homozygotes.

<sup>d</sup> The over-dominant model is defined as the major allele homozygotes + minor allele homozygotes vs. heterozygotes.

fold increase in the risk of OLP in the recessive model (OR = 3.95, 95% CI = 0.79–21.69,  $P = 0.05$ ) (TT vs. CC+CT genotypes). Also, in the over-dominant model, the CT genotype was related to a 1.78-fold higher risk of OLP (OR = 1.78, 95% CI = 1.01–3.12,  $P = 0.03$ ) (Table 2). This shows that *H19* rs217727 polymorphism positively correlates with the risk of OLP. On the other hand, age, gender, and type of OLP were not significantly related to *H19* rs217727 polymorphism.

## DISCUSSION

This study assessed *H19* gene polymorphism (rs217727) in OLP patients compared with controls. We applied inheritance models in multiple forms to assess the relationship between SNPs in the *H19* gene and OLP risk. A relation was found between OLP and the *H19* rs217727 polymorphism in dominant, codominant, and over-dominant models. However, age, gender, and type of OLP were not significantly related to *H19* rs217727 polymorphism. Regarding the recessive inheritance model, we found no relationship between SNPs and the risk of OLP. Our findings indicate a new diagnostic marker for OLP, possibly reflecting a higher predisposition to this condition. It may also be a criterion for cancer susceptibility in OLP patients, and *H19* gene polymorphisms might discriminate between potentially malignant and non-harmful lesions.

Oral lichen planus (OLP) is the most common

oral lesion with a possible malignant change. The etiopathogenesis remains elusive, though it must be promptly diagnosed to facilitate effective care and diminish the seriousness of its influence on the quality of life. Hence, delineating the exact cellular and molecular mechanisms causing OLP development to OSCC is essential<sup>23-25</sup>. Presently, tissue biopsy and histological evaluation is a gold and ideal technique for diagnosing OLP and other malignant and premalignant oral mucosa lesions, but it is costly and may pose risks. Hence, non-invasive techniques, such as assessing certain biomarkers, have been established as alternatives<sup>3,23,26</sup>.

Various components, comprising inflammatory cells, cytokines, tumor suppressor genes, and matrix metalloproteinases, can induce OLP advancement. Numerous surveys have concentrated on proteins that control the immune system and autoimmunity<sup>3,7,27-30</sup>. We found that the polymorphism of lncRNA *H19* significantly correlated with OLP susceptibility in all forms of the disorder compared to healthy subjects. Because the molecular alterations happen before the morphological variations in precancerous and cancerous cells, lncRNA *H19* may give important evidence on patient prognosis, follow-up, and management.

Long non-coding RNAs (lncRNAs) are supervisory layers in transcriptional and post-transcriptional gene control<sup>13,14,31</sup>. The expression stage, composition, and strength of lncRNA may be altered by a mutation

in the lncRNA, thus leading to the development of several diseases<sup>18,20</sup>. Several studies show that lncRNAs represent an important biomarker in patients with head and neck cancers<sup>15,20,21,32-34</sup>. Jia *et al.* investigated the effect of lncRNAs on the pathogenesis, diagnosis, and prognosis of cancerous changes from dysplasia to SCC in the oral mucosa. They showed that lncRNAs were significantly expressed in both SCC and dysplastic states; the target genes possibly have a crucial effect on the carcinogenesis and expansion of oral cancers<sup>14</sup>.

Numerous studies have confirmed the considerable effect of lncRNA *H19* on various tumor cells and carcinogenesis<sup>16,19-21</sup>. Our previous study found that lncRNA *H19* rs217727 was statistically related to OSCC susceptibility in an Iranian population<sup>20</sup>. A study indicated that *H19* may improve gastric tumor cell invasion and migration<sup>35</sup>. Elias *et al.* reported that lncRNA *H19* might be a useful diagnostic biomarker for breast tumors. *H19* could act as a valuable marker classifying the specimens in typical vs. benign vs. premalignant breast tissues and help to determine aggressive from nonaggressive lesions<sup>36</sup>. A high serum level of *H19* was found in cases with certain myelomas and non-small cell lung cancers, and this biomarker appears useful for the early detection and prognosis of such tumors<sup>37-39</sup>.

In known cases of malignancy, *H19* represents a predictive factor. Reduced levels of *H19* in tumor tissue can suggest aggressive characteristics, meaning that *H19* levels can influence treatment planning<sup>36</sup>. Some researchers have described the variation in *H19* levels found in tissues<sup>11,40</sup>. Chu and colleagues described the relationship between *H19* polymorphisms and tumorigenicity<sup>41</sup>. Other studies considered the association linking cancer predisposition and lncRNA *H19* polymorphisms<sup>42,43</sup>. In one study, the rs2839698 G > A polymorphism was accompanied by gastrointestinal tract malignancy<sup>43</sup>. This is in line with our study, which linked the rs217727 C allele with a reduced risk of OLP.

Huang *et al.* assessed the association of the polymorphisms of *H19* (rs2839698, rs3741219) with autoimmune disease in Chinese individuals. They found no significant correlation between these diseases and *H19* polymorphism in the Han population<sup>44</sup>. This does not align with our work, which showed a significant association of *H19* rs217727

polymorphism with OLP as an autoimmune disease in an Iranian population. This discrepancy may be related to differences in study populations and sample sizes. It has been noted that the expression of *H19* is associated with hormone receptors<sup>45</sup>; most of our patients who suffered from OLP were females, possibly correlating with a higher serum level of *H19*.

A study showed that the T allele of rs217727 was more frequently seen in type 2 diabetes in an Iranian population than in healthy controls. Likewise, the rs217727-TT genotype was highly correlated with type 2 diabetes<sup>46</sup>. This finding is in accordance with our study, which showed that the prevalence of the TT genotype was significantly greater in OLP patients compared to controls (5.71% vs 1.5%).

Previous studies showed that lncRNA can dysregulate immune responses and harbor pro-inflammatory cytokines<sup>47-49</sup>. Extensive data indicate that immunologic and inflammatory factors have significant responsibilities in OLP. Furthermore, microRNAs (miRs), small non-coding RNAs, are implicated in OLP. Specifically, miR-155 is upregulated in patients with OLP; miR-155 has various roles and is strongly linked to inflammation and immune system regulation<sup>50</sup>.

It is important to note that while earlier surveys have described the role of *H19* in a variety of disorders<sup>51</sup>, this investigation is the earliest to examine the genetic variation of this gene in relationship with OLP. Our study assessed the serum samples of patients with confirmed OLP. Nonetheless, when talking about limitations, due to COVID-19 pandemic our sample size was chosen as the least size calculated by previous studies. Further studies with larger sample sizes are suggested. Other constraints of the current research were the diverse ethnic population that lives in this region. Further research on single ethnic groups are also suggested.

## CONCLUSION

In conclusion, the potential premalignant characteristic of OLP highlights the importance of investigating the underlying mechanism of this disorder. A reliable and objective molecular marker is required to assist clinicians in making appropriate clinical judgments concerning high-risk disorders without undertaking tissue biopsy or alongside the histopathological findings. The present study sought

to address this issue, demonstrating a significant link between lncRNA *H19* polymorphism and OLP lesions. However, our investigation is just the beginning, and further studies are needed to delineate the potential roles of lncRNAs in OLP.

### Authors contributions

JG, MZ, MJM, MJF and FR conceived and designed the research and experiments; MJM, MJF and NM Performed the experiments; NM and MZ Analyzed and interpreted the data; MJM, JG and FR wrote the draft of the manuscript. MZ performed the editing and the final manuscript was read and confirmed by all the authors.

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**Conflict of Interest:** None declared.

## REFERENCES

- Eisen D. The clinical features, malignant potential, and systemic associations of oral lichen planus: a study of 723 patients. *J Am Acad Dermatol.* 2002;46(2):207-14.
- Moshaverinia M, Rezazadeh F, Dalvand F, et al. The relationship between oral lichen planus and blood group antigens. *World J Med Sci.* 2014;10(2):103-5.
- Carrozzo M, Porter S, Mercadante V, et al. Oral lichen planus: a disease or a spectrum of tissue reactions? types, causes, diagnostic algorithms, prognosis, management strategies. *Periodontology.* 2019;80(1):105-25.
- Ghapanchi J, Andisheh-Tadmir A, Torkaman P, et al. Evaluation of the serum levels of galectin-3 in patients with oral lichen planus disease. *Oral Dis.* 2019;25(2):466-70.
- González-Moles MÁ, Warnakulasuriya S, González-Ruiz I, et al. Worldwide prevalence of oral lichen planus: a systematic review and meta-analysis. *Oral Dis.* 2021;27(4):813-828.
- Amanat D, Ebrahimi H, Zahedani MZ, et al. Comparing the effects of cryotherapy with nitrous oxide gas versus topical corticosteroids in the treatment of oral lichen planus. *Indian J Dent Res.* 2014;25(6):711-6.
- Ghapanchi J, Ghaderi H, Haghshenas MR, et al. Observational molecular case-control study of genetic polymorphisms 1 in programmed cell death protein-1 in patients with oral lichen planus. *Asian Pac J Cancer Prev.* 2019;20(2):421-4.
- González-Moles MÁ, Ruiz-Avila I, Gonzalez-Ruiz L, et al. Malignant transformation risk of oral lichen planus: a systematic review and comprehensive meta-analysis. *Oral Oncol.* 2019;96:121-30.
- Idrees M, Kujan O, Shearston K, et al. Oral lichen planus has a very low malignant transformation rate: a systematic review and meta-analysis using strict diagnostic and inclusion criteria. *J Oral Pathol Med.* 2021;50(3):287-98.
- Ghapanchi J, Haghshenas MR, Ghaderi H, et al. Ctlα-4 gene polymorphism in +49 a/g position: a case control study on patients with oral lichen planus. *J Int Oral Health.* 2014;6(5):17-21.
- Gibb EA, Enfield KS, Stewart GL, et al. Long non-coding RNAs are expressed in oral mucosa and altered in oral premalignant lesions. *Oral Oncol.* 2011;47(11):1055-61.
- Dhanoa JK, Sethi RS, Verma R, et al. Long non-coding RNA: its evolutionary relics and biological implications in mammals: a review. *J Anim Sci Technol.* 2018;60(1):1-10.
- Jia H, Wang X, Sun Z. Exploring the molecular pathogenesis and biomarkers of high risk oral premalignant lesions on the basis of long noncoding RNA expression profiling by serial analysis of gene expression. *Eur J Cancer Prev.* 2018;27(4):370.
- Jia H, Wang X, Sun Z. Exploring the long noncoding RNAs-based biomarkers and pathogenesis of malignant transformation from dysplasia to oral squamous cell carcinoma by bioinformatics method. *Eur J Cancer Prev.* 2020;29(2):174.
- Jia H, Wang X, Sun Z. Screening and validation of plasma long non-coding RNAs as biomarkers for the early diagnosis and staging of oral squamous cell carcinoma. *Oncol Lett.* 2021;21(2):1.
- Lu Y, Tan L, Shen N, et al. Association of lncRNA *H19* rs217727 polymorphism and cancer risk in the Chinese population: a meta-analysis. *Oncotarget.* 2016;7(37):59580-8.
- Leighton PA, Ingram RS, Eggenschwiler J, et al. Disruption of imprinting caused by deletion of the *H19* gene region in mice. *Nature.* 1995;375(6526):34-9.
- Harati-Sadegh M, Kohan L, Teimoori B, et al. The long non-coding RNA *H19* rs217727 polymorphism is associated with PE susceptibility. *J Cell Biochem.* 2018;119(7):5473-80.
- Yoshimizu T, Miroglio A, Ripoche MA, et al. The *H19* locus acts in vivo as a tumor suppressor. *Proc Natl Acad Sci.* 2008;105(34):12417-22.
- Ghapanchi J, Ranjbar Z, Mokhtari MJ, et al. The lncRNA *H19* rs217727 polymorphism is associated with oral squamous cell carcinoma susceptibility in Iranian population. *BioMed Res Int.* 2020;2020:1634252.
- Ghapanchi J, Mokhtari MJ, Zahed M, et al. Genetic analysis of lncRNA *H19* (rs217727) and *MIAT* (rs1894720) polymorphisms in patients with salivary gland tumors.

- Gene Rep. 2023;30:101724.
22. Guo Q, Wang H, Wang Y. LncRNA H19 polymorphisms associated with the risk of OSCC in Chinese population. *Eur Rev Med Pharmacol Sci.* 2017;21:3770-4.
  23. Muniz JM, Borges CRB, Beghini M, et al. Galectin-9 as an important marker in the differential diagnosis between oral squamous cell carcinoma, oral leukoplakia and oral lichen planus. *Immunobiol.* 2015;220(8):1006-11.
  24. Rezazadeh F, Shahbazi F, Andisheh-Tadbeer A. Evaluation of salivary level of IL-10 in patients with oral lichen planus, a preliminary investigation. *Comp Clin Path.* 2017;26(3):531-4.
  25. Zahed M, Goli A, Zamirian E, et al. Spatial evaluation of oral lichen planus patients referred to Shiraz dental school, Iran: a medical geography approach. *J Dent.* 2022;23:361-8.
  26. Ranjbar MA, Ranjbar Z, Zahed M, et al. CD73 a novel marker for the diagnosis of benign and malignant salivary gland tumors. *J Clin Exp Dent.* 2019;11(3):e213.
  27. Ghabanchi J, Fattahi MJ, Mardani M, et al. Polymorphism of tumor protein P53 codon 72 showed no association with oral lichen planus in Shiraz, Iran. *J Craniofac Surg.* 2009;20(6):2168-70.
  28. Gholijani N, Daryabor G, Kalantar K, et al. Interleukin-27 gene variant rs153109 is associated with enhanced cytokine serum levels and susceptibility to Behçet's disease in the Iranian population. *Eur Cytokine Netw.* 2020;31(4):140-6.
  29. Gholijani N, Daryabor G, Yazdani MR, et al. Serum interleukin-37 (IL-37) and its gene polymorphism in Iranian Behçet's disease patients: association with disease manifestations and activity. *Meta Gene.* 2020;26:100794.
  30. Lavaee F, Ghapanchi J, Anjomruz A, et al. The evaluation of the serum level of IL-10 in OLP patients. *Comp Clin Path.* 2018;27(1):131-4.
  31. Yang Z, Li X, Yang Y, et al. Long noncoding RNAs in the progression, metastasis, and prognosis of osteosarcoma. *Cell Death Dis.* 2016;7(9):e2389.
  32. Feng L, Chen W, Qiu W. Long non-coding RNAs associated with oral squamous cell carcinoma. *Eur Rev Med Pharmacol Sci.* 2019;23(20):8888-96.
  33. Cossu AM, Mosca L, Zappavigna S, et al. Long non-coding RNAs as important biomarkers in laryngeal cancer and other head and neck tumours. *Int J Mol Sci.* 2019;20(14):3444.
  34. Wu H, Zheng J, Deng J, et al. A genetic polymorphism in lincRNA-uc003opf. 1 is associated with susceptibility to esophageal squamous cell carcinoma in Chinese populations. *Carcinogenesis.* 2013;34(12):2908-17.
  35. Chen J, Wang Y, Zhang X, et al. H19 serves as a diagnostic biomarker and up-regulation of H19 expression contributes to poor prognosis in patients with gastric cancer. *Neoplasma.* 2016;63(2):223-30.
  36. Elias-Rizk T, El Hajj J, Segal-Bendirdjian E, et al. The long non coding RNA H19 as a biomarker for breast cancer diagnosis in Lebanese women. *Sci Rep.* 2020;10(1):22228.
  37. Luo J, Li Q, Pan J, et al. Expression level of long noncoding RNA H19 in plasma of patients with nonsmall cell lung cancer and its clinical significance. *J Cancer Res Ther.* 2018;14(4):860.
  38. Pan Y, Chen H, Shen X, et al. Serum level of long noncoding RNA H19 as a diagnostic biomarker of multiple myeloma. *Clin Chim Acta.* 2018;480:199-205.
  39. Lv J, Ma L, Chen XL, et al. Downregulation of LncRNAH19 and MiR-675 promotes migration and invasion of human hepatocellular carcinoma cells through AKT/GSK-3 $\beta$ /Cdc25A signaling pathway. *J Huazhong Univ Sci Technol Med Sci J.* 2014;34(3):363-9.
  40. Dugimont T, Curgy JJ, Wernert N, et al. The H19 gene is expressed within both epithelial and stromal components of human invasive adenocarcinomas. *Biol Cell.* 1995;85(2-3):117-24.
  41. Chu M, Yuan W, Wu S, et al. Quantitative assessment of polymorphisms in H19 lncRNA and cancer risk: a meta-analysis of 13,392 cases and 18,893 controls. *Oncotarget.* 2016;7(48):78631.
  42. Yang PJ, Hsieh MJ, Hung TW, et al. Effects of long noncoding RNA H19 polymorphisms on urothelial cell carcinoma development. *Int J Environ Res Public Health.* 2019;16(8):1322.
  43. Li XF, Yin XH, Cai JW, et al. Significant association between lncRNA H19 polymorphisms and cancer susceptibility: a meta-analysis. *Oncotarget.* 2017;8(28):45143.
  44. Huang AF, Su LC, Jia H, et al. No association of single nucleotide polymorphisms within H19 and HOX transcript antisense RNA (HOTAIR) with genetic susceptibility to systemic lupus erythematosus, rheumatoid arthritis, and primary Sjögren's syndrome in a Chinese Han population. *Clin Rheumatol.* 2017;36(11):2447-53.
  45. Sun H, Wang G, Peng Y, et al. H19 lncRNA mediates 17 $\beta$ -estradiol-induced cell proliferation in MCF-7 breast cancer cells. *Oncol Rep.* 2015;33(6):3045-52.
  46. Ghaedi H, Zare A, Omrani MD, et al. Genetic variants in long noncoding RNA H19 and MEG3 confer risk of type 2 diabetes in an Iranian population. *Gene.* 2018;675:265-71.
  47. Wang X, Sun W, Shen W, et al. Long non-coding RNA DILC regulates liver cancer stem cells via IL-6/STAT3 axis. *J Hepatol.* 2016;64(6):1283-94.
  48. Han Y, Kang C, Kang M, et al. Long non-coding RNA Mirt2 prevents TNF- $\alpha$ -triggered inflammation via the repression of microRNA-101. *Int Immunopharmacol.* 2019;76:105878.
  49. Hadjicharalambous MR, Roux BT, Feghali-Bostwick CA, et al. Long non-coding RNAs are central regulators of the IL-1 $\beta$ -induced inflammatory response in normal and idiopathic pulmonary lung fibroblasts. *Front Immunol.* 2018;9:2906.
  50. Tao Y, Ai R, Hao Y, et al. Role of miR-155 in immune regulation and its relevance in oral lichen planus. *Exp Ther Med.* 2019;17(1):575-86.
  51. Ghapanchi J, Mokhtari MJ, Zahed M, Ardekani ST, Fattahi MJ, Khademi B, Asadabadi T, Koohpeima F, Arab S, Avandi S, Namvaran MR. Genetic analysis of lncRNA H19 (rs217727) and MIAT (rs1894720) polymorphisms in patients with salivary gland tumors. *Gene Reports.* 2023 Mar 1;30:101724.